

## Human microRNAs profiling in response to influenza A viruses (subtypes pH1N1, H3N2, and H5N1)

Jarika Makkoch<sup>1</sup>, Witthaya Poomipak<sup>2</sup>, Suthat Saengchoowong<sup>3</sup>, Kritsada Khongnomnan<sup>1</sup>, Kesmanee Praianantathavorn<sup>1</sup>, Thananya Jinato<sup>4</sup>, Yong Poovorawan<sup>5</sup> and Sunchai Payungporn<sup>1,4</sup>

<sup>1</sup>Department of Biochemistry, Faculty of Medicine, Chulalongkorn University, Bangkok 10330 Thailand; <sup>2</sup>Research affairs, Faculty of Medicine, Chulalongkorn University, Bangkok 10330 Thailand; <sup>3</sup>Joint Chulalongkorn University – University of Liverpool PhD Programme in Biomedical Sciences and Biotechnology, Bangkok 10330, Thailand; <sup>4</sup>Systems Biology Center, Research Affairs, Faculty of Medicine, Chulalongkorn University, Bangkok, 10330 Thailand; <sup>5</sup>Center of Excellence in Clinical Virology, Faculty of Medicine, Chulalongkorn University, Bangkok 10330, Thailand

Corresponding author: Sunchai Payungporn. Email: sp.medbiochemcu@gmail.com

### Abstract

MicroRNAs (miRNAs) play an important role in regulation of gene silencing and are involved in many cellular processes including inhibition of infected viral replication. This study investigated cellular miRNA expression profiles operating in response to influenza virus in early stage of infection which might be useful for understanding and control of viral infection. A549 cells were infected with different subtypes of influenza virus (pH1N1, H3N2 and H5N1). After 24 h post-infection, miRNAs were extracted and then used for DNA library construction. All DNA libraries with different indexes were pooled together with equal concentration, followed by high-throughput sequencing based on MiSeq platform. The miRNAs were identified and counted from sequencing data by using MiSeq reporter software. The miRNAs expressions were classified into up and downregulated miRNAs compared to those found in non-infected cells. Mostly, each subtype of influenza A virus triggered the upregulated responses in miRNA expression profiles. Hsa-miR-101, hsa-miR-193b, hsa-miR-23b, and hsa-miR-30e\* were upregulated when infected with all three subtypes of influenza A virus. Target prediction results showed that virus infection can trigger genes in cellular process, metabolic process, developmental process and biological regulation. This study provided some insights into the cellular miRNA profiling in response to various subtypes of influenza A viruses in circulation and which have caused outbreaks in human population. The regulated miRNAs might be involved in virus–host interaction or host defense mechanism, which should be investigated for effective antiviral therapeutic interventions.

**Keywords:** miRNAs, influenza A virus, expression profiles, next-generation sequencing

*Experimental Biology and Medicine* 2016; 241: 409–420. DOI: 10.1177/1535370215611764

### Introduction

The pandemic influenza virus usually infects a large population, not only human but also livestock, such as swine and poultry, resulting in problems in public health and loss of economic distress in terms of hospitalization.<sup>1,2</sup> More than 284,000 people worldwide died because of pH1N1 after a year of pandemic.<sup>3</sup> Some strains of influenza virus can be zoonotic, such as H5N1, leading to great losses in the poultry industry, and with high mortality rate in reported human cases (>60%).<sup>4–8</sup> Unlike being infected with seasonal influenza, H5N1 infection can cause multi-organ failure, hemorrhagic syndrome and cytokine storm, suggesting that host genes can respond in a different manner depending on the specific strain of influenza virus infection. Previous studies showed that influenza virus principally controls host genes to facilitate viral replication in multiple

steps, and one of the potential mechanisms to control gene expression is microRNA (miRNA) regulation.<sup>9–13</sup>

MiRNAs are small (17–24 nt in length), endogenous, non-coding RNAs found in multicellular organisms.<sup>14</sup> They are acknowledged as vigorous tool regulator molecules that can trigger the targeted gene functions by the translational repression or mRNA degradation through the RNA interference pathway.<sup>14–18</sup> Previous studies showed that the genome of more than 150 species including viruses contain sequences of mature miRNAs, especially in the human genome, in which more than 1000 miRNAs have been characterized.<sup>16,19–21</sup> Studies have expressed that a broad range of cellular mechanisms are mediated by miRNAs, such as cell proliferation signaling, chromosome maintenance, cell differentiation and apoptosis.<sup>9,14,22–24</sup> Not only can viruses regulate host gene expression by using miRNAs to promote

the viral life cycle, but the host cell can also alter differential gene expression to respond against viral infection via the miRNA mechanism. The proportion of the miRNA level due to the encounter between host and virus will indicate the disease progression and outcome.<sup>25,26</sup> Multiple pieces of evidence have shown that both host and virus constructed miRNAs are essential for virus propagation. It has been reported that the influenza virus plays a critical role in the modulation of the miRNA mechanisms of infected cells and animal models.<sup>9,27-29</sup> In contrast, miRNAs can also react with the influenza virus replication process. Many previous studies have mentioned that influenza virus replication and propagation were inhibited due to cellular miR-323, miR-491, miR-654 and miR-146a expression which specifically bind to the PB1 gene of the virus.<sup>30,31</sup>

Many techniques have been used to validate the miRNA screening and target prediction of miRNA response to influenza virus infection, such as qPCR, luciferase assay and drug compound library screening.<sup>9,12,28,31</sup> Among these, microarray techniques have been used extensively to study the miRNA profiles in response to the pathogenesis of the influenza virus.<sup>32-35</sup> Although microarray technique can gain a large amount of data of miRNA expression, it cannot determine the significance of particular genes in the cellular mechanism which are not covered by the probe set, and may lead to some essential genes, which slightly change, being dismissed. The high-throughput next-generation sequencing (NGS) can cope with these problems, as the dataset generated from NGS will demonstrate the genome-wide miRNA expression profiles of infected host cells. This study aimed to investigate cellular miRNA expression profiles operating in response to three specific strains of influenza A virus responsible for causing outbreaks in the human population; Avian-origin highly pathogenic avian influenza (HPAI) H5N1, human pandemic (pH1N1) and seasonal H3N2 influenza A virus infection. The study also aimed to predict target genes which might be useful for understanding the host defense mechanism in terms of regulating viral infection.

## Materials and methods

### Cell culture

A549 (human lung carcinoma) cells used in this study were kindly provided by Prof. Jen-Ren Wang, Department of Medical Laboratory Science and Biotechnology, Center of Infectious Disease and Signaling Research, National Cheng Kung University. A549 cells were cultured in Dulbecco's modified eagle medium (DMEM) (Thermo Scientific) containing 10% heat inactivated fetal bovine serum (Thermo Scientific) and 1% (v/v) antibiotic/antimycotic (Gibco) under humidified atmosphere with 5% CO<sub>2</sub> at 37°C.

### Virus infection

A549 cells were seeded ( $3 \times 10^5$  cells/well) in DMEM medium without antibiotic/antimycotic in 6-well plates. Then influenza A viruses subtype pH1N1 (A/Thailand/

104/2009), H3N2 (A/Thailand/CU-H187/2010) and H5N1 (A/Thailand/NK165/2005) were prepared by diluting the virus stocks with DMEM medium to yield the desired concentration of  $3 \times 10^5$  pfu/mL (MOI=0.5). Each viral infection was performed in triplicate in a biosafety level 3 (BSL3) laboratory, Faculty of Medicine, Chulalongkorn University. When cells reached approximately 80% confluence, cell culture media were removed from each well and cells were washed with 3 mL of phosphate buffer saline (PBS). After that 500 µL of each virus suspension was added into each well and then incubated in a humidified atmosphere with 5% CO<sub>2</sub> at 37°C for 1 h with occasional shaking to allow the virus to adsorb into the cells. After incubation, the virus suspension was removed from each well and then washed by 3 mL of PBS. Finally, 3 mL of complete DMEM medium was added into each well and cells were incubated for 24 h. The protocol of this study was approved by the Institutional Review Board (IRB No. 121/55), Faculty of Medicine, Chulalongkorn University.

### Library preparation

After 24 h post infection, triplicate wells were pooled together and microRNAs were extracted by using a microRNA purification kit (Norgen) following the manufacturer's protocol. The concentrations of microRNA were quantified by using Qubit fluorometer (Invitrogen) with a Quant-iT<sup>TM</sup> RNA Assay kit (Invitrogen). Purified microRNA (0.2 µg) from each group (uninfected cells, pH1N1 infected cells, H3N2 infected cells and H5N1 infected cells) were used to construct four libraries with different indexes according to a TruSeq Small RNA sample preparation kit (Illumina). The length of DNA libraries were validated by using Agilent 2100 Bioanalyzer system (Agilent) with a high sensitivity DNA chip (Agilent). The concentrations of DNA libraries were quantified by using Qubit fluorometer (Invitrogen) with a Quant-iT<sup>TM</sup> DNA Assay kit (Invitrogen). The four DNA libraries were pooled together with equal concentration and then high-throughput sequenced using a MiSeq platform (Illumina).

### Analysis of microRNA expression profiles

MiSeq reporter software version 2.4 was used for analysis of sequencing data. Low-quality reads (Q-score < 30) were excluded whereas low-quality regions of sequences were trimmed. The passing filtered reads (Q-score ≥ 30) were aligned with human genomic DNA (hg19), mature & precursor human miRNA (from miRbase) and contaminant RNA (human tRNA, rRNA and mRNA) using the Bowtie algorithm (<http://bowtiebio.sourceforge.net>). The clusters which matched to the miRNA database but not contaminant RNA were considered as miRNA. The miRNAs were classified and counted based on the number of reads matched to the miRbase ([www.mirbase.org/](http://www.mirbase.org/)).

## Differential expression analysis and data visualization

The data of validated miRNAs expressed in cell culture obtained from next-generation sequencing platform were demonstrated in form of fold change, compared to the amount of those miRNAs expressed in control experiment (mock infection). Differential expression analysis was calculated in terms of fold changes

$$\text{Fold changes} = \log_2 \left( \frac{\text{Normalized expression in case}}{\text{Normalized expression in control}} \right)$$

$$\begin{aligned} &\text{Normalized expression in control} \\ &= \frac{\text{miRNA of interest in control}}{\text{Total count of miRNAs in control}} \end{aligned}$$

$$\begin{aligned} &\text{Normalized expression in case} \\ &= \frac{\text{miRNA of interest in case}}{\text{Total count of miRNAs in case}} \end{aligned}$$

The data visualization was performed by using the GraphPad Prism 5 software to demonstrate the up-regulated and deregulated miRNAs.

## Analysis of microRNA-target genes and biological process classification

Identifications of target genes for selected miRNAs were analyzed by using the experimentally validated miRNA-target interactions data accumulated in miRTarBase version 4.5 (<http://mirtarbase.mbc.nctu.edu.tw/>). Briefly, the target genes of miRNAs were selected based on strong experimental evidence obtained from validation methods including reporter assay, qPCR or western blot. The PANTHER (Protein ANalysis THrough Evolutionary Relationships) classification system version 9.0 (<http://www.pantherdb.org/>)<sup>36</sup> was used to classify genes and proteins in order to facilitate high-throughput analysis. Each of the validated target genes of miRNA were classified based on biological processes including apoptotic process (GO:0006915), biological adhesion (GO:0022610), biogenesis (GO:0071840), biological regulation (GO:0065007), cellular process (GO:0009987), developmental process (GO:0032502), immune system process (GO:0002378), localization (GO:0051179), multicellular organismal process (GO:0032501), metabolic process (GO:0008152), reproduction (GO:0000003), and response to stimulus (GO:0050896).

## Prediction of protein-protein interaction

The target genes of significant miRNAs from NGS experiment were performed for protein interaction. The interactions among the proteins in this study were analyzed from the STRING 9.1 database,<sup>37,38</sup> which merges and weighs information from various sources, covering conserved neighborhood, gene fusions, phylogenetic occurrence, co-expression, experiments, database imports, and text-mining.<sup>39</sup> The scores greater than 0.7 are considered as the high confidence.

## Results

### Analysis of human miRNAs by NGS based on MiSeq platform

After 24 h of infection, triplicate wells of each group were pooled together in order to generate 4 DNA libraries including uninfected control, pH1N1, H3N2 and H5N1 influenza A virus infected cells. Then the 4 libraries with different indexes were subjected to high-throughput sequencing using MiSeq platform. A total of 3,654,615 reads were obtained from a high-throughput sequencing process. After data filtering based on quality score ( $Q \geq 30$ ), 2,225,781 high-quality filtered reads were obtained and used for further analysis. The overall reads with a perfect match compared to sequences of miRNA database (miRBase version 16.0) ([www.mirbase.org/](http://www.mirbase.org/)) were characterized and counted. According to miRBase data, the results showed that there were totally 19,732 reads belonging to human miRNAs, including 4604, 6482, 4509 and 4137 of identified reads which were obtained from non-infected, pH1N1-infected, H3N2-infected and H5N1-infected cells, respectively.

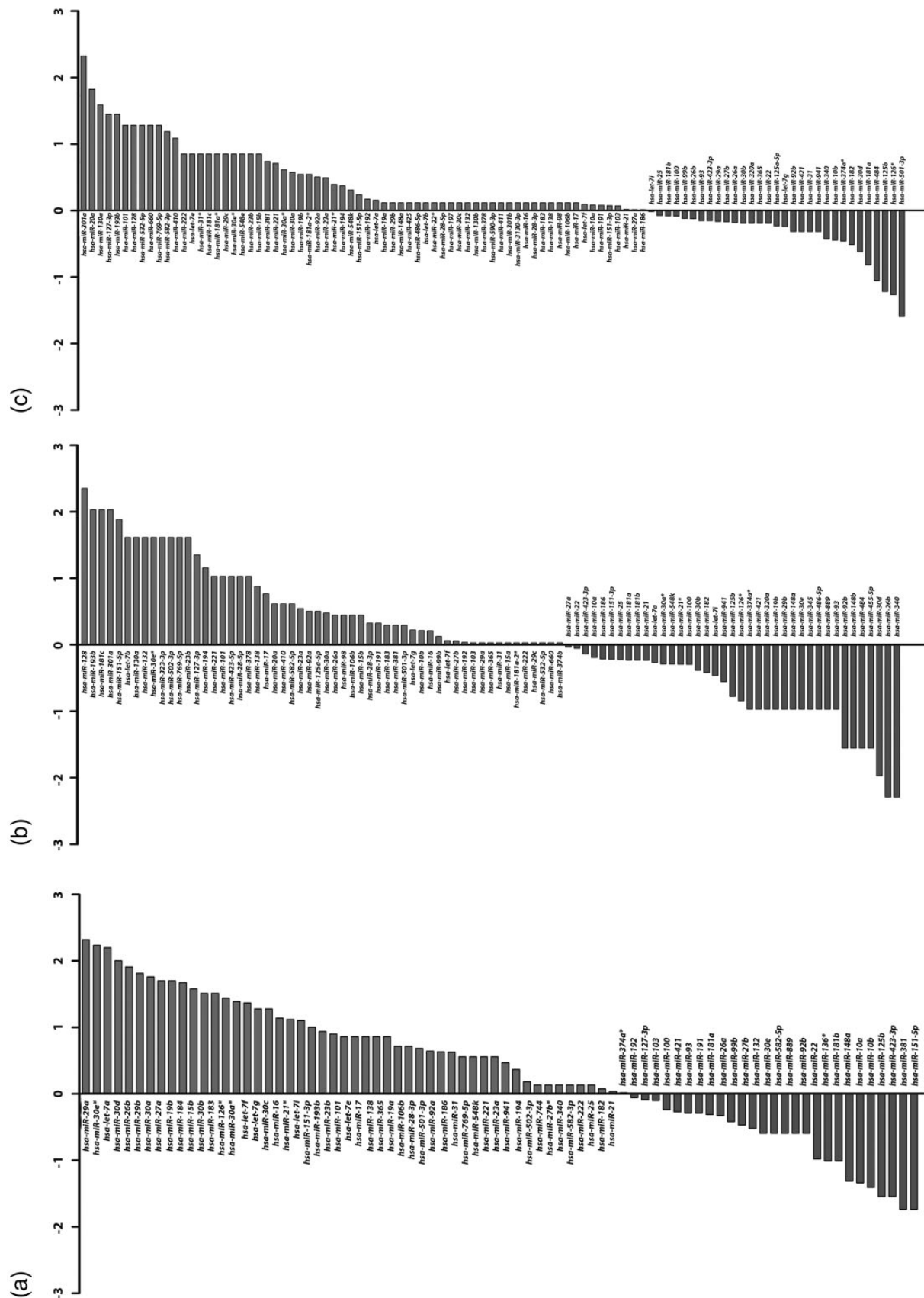
### Differential expression of miRNA profiling

MiRNAs expression profiles of A549 cells infected with each subtype of influenza A virus were compared with uninfected cells. The list of differentially expressed miRNAs was identified and classified as upregulated ( $>1.5$  fold increase) and downregulated ( $<1.5$  fold decrease) compared to those miRNA levels in uninfected cells. The full list of miRNA profiling is shown in Figure 1. A549 infected with pH1N1 influenza A virus expressed 10 downregulated miRNAs and 29 upregulated miRNAs. Infection of H3N2 influenza A virus showed seven downregulated miRNAs and 20 upregulated miRNAs while A549 cells infected with HPAI H5N1 influenza virus revealed five downregulated and 23 upregulated miRNAs. The miRNA expression profiles of cells infected with each subtype of influenza A virus were compared both in upregulated (Figure 2a) and downregulated (Figure 2b) miRNAs in order to identify common miRNA responses to multiple subtypes of influenza A virus. The results showed that four miRNAs including hsa-miR-101, hsa-miR-193b, hsa-miR-23b and hsa-miR-30e\* were found to have upregulated expression when infected with all three strains of influenza A virus. The hsa-miR-128, hsa-miR-130a, hsa-miR-181c, hsa-miR-301a, hsa-miR-127-3p and hsa-miR-769-5p were upregulated in cells infected with H3N2 and H5N1 while let-7e and hsa-miR-15b were upregulated by pH1N1 and H5N1 infection. For downregulated miRNAs, hsa-miR-125b was downregulated by H5N1 and pH1N1 infection while only hsa-miR-484 was downregulated by H5N1 and H3N2 influenza viruses.

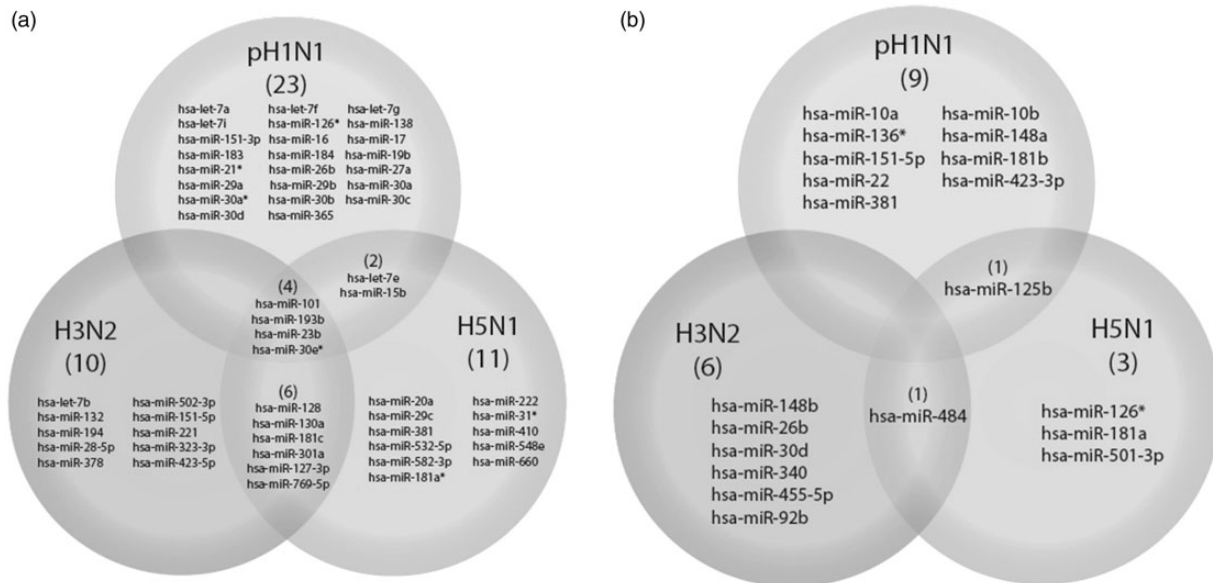
### Analysis of common miRNA profiles in response to influenza A virus infection

MiRNA expression profiles obtained from this study were compared to previous studies which were performed in various subtypes of influenza A virus and cellular





**Figure 1** The miRNA expression profiles in each subtypes of influenza A virus infection. A549 cells were infected (MOI = 0.5) with three subtypes of influenza virus. (a) Seventy-eight miRNAs were expressed after infected with pH1N1 influenza virus, including 10 downregulated miRNAs and 39 miRNAs that expressed in the similar level as control cells. (b) Infection of H3N2 influenza virus showed total of 95 miRNAs expressed, including seven downregulated miRNAs and 68 miRNAs that expressed in the similar level as control cells. (c) Ninety-nine miRNAs were responded in A549 cells infected with HPAI H5N1 influenza virus, five of which were downregulated, 23 miRNAs were expressed similar level compared to uninfected control



**Figure 2** Venn-Euler diagram shows a summary of differentially expressed miRNAs in response to influenza A virus infection. (a) Upregulated miRNAs when infected with pH1N1, H5N1 and H3N2. (b) Downregulated miRNAs when the cells were infected by pH1N1, H3N2, and H5N1

models.<sup>12,28,40–45</sup> The comparative data suggested that there were 15 common miRNAs including let-7 family, miR-10, miR-15, miR-21, miR-29, miR-30, miR-34, miR-101, miR-132, miR-146, miR-148, miR-548, and miR-574 (Table 1).

### Host target genes prediction of miRNA expression profile

After the analysis of common miRNA profiles in response to influenza A virus infection compared to several studies, 15 miRNAs were selected to be significant miRNAs that responded to influenza A virus infection. This study examined the target genes of miRNAs by miRNA-target interaction database called miRTarBase version 4.5 (<http://mirtarbase.mbc.nctu.edu>) and the PANTHER classification system. Using the information in the database for target prediction, the results showed that each miRNA could interact with hundreds of genes. We narrowed the targeted genes of miRNAs from the profile using strong experimental evidence of validation methods from previous studies. Using the PANTHER system, the number of target genes of the 15 selected miRNAs were summarized (Table 2). The number and percentages of miRNA target genes based on GO biological process classification of selected target genes were demonstrated. Each of the validated target genes for miRNAs was categorized based on types of biological processes, including apoptotic process (AP), biological adhesion (BA), biogenesis (BG), biological regulation (BR), cellular process (CP), developmental process (DP), immune system process (IP), localization (Lo), multicellular organismal process (MO), metabolic process (MP), reproduction (Re), and response to stimulus (Rs) (Table S1).

The protein-protein interactions provide information regarding most biological processes in an organism. In this study, the prediction of target proteins interaction via four deregulated miRNAs from our experiment, and

therefore miR-101, miR-193b, miR-23b, and miR-30a, were illustrated. The analysis of topology demonstrates important network illustrations created by interacting proteins. Therefore, the interactions between proteins from the STRING 9.1 database were used to form the target gene network of the differentially expressed microRNAs to determine several 'hub' nodes (Figure 3), which play an important role in apoptosis during the infection of influenza viruses. This finding may enlighten the understanding of the promising roles of the differentially expressed microRNAs.

### Viral target genes prediction of miRNAs

Many previous studies showed that the cellular miRNAs can also target and modulate viral genes.<sup>11,46</sup> In this study, both the downregulated and upregulated miRNAs underwent the influenza virus target prediction compared to previous studies.<sup>30,47,48</sup> The result indicated that out of 11 miRNAs, nine of them were upregulated and targeted influenza virus genes. In this group, some of them can target only one viral gene, such as in the cases of miR-127, 128 and 29a, which target only PB1, PA, and HA gene, respectively, while some miRNAs, including miR-548, 222, and 16 can have multiple gene targets (Table 3). Most of the viral genes targeted by miRNAs in this study were related to polymerase activity (PB2, PB1 and PA).

### Discussion

The present study provided miRNA profiling at an early stage of infection with three subtypes of influenza A virus (pH1N1, H3N2 and H5N1) by using next-generation sequencing. From the miRNA profiling results it was found that a number of host genes could be regulated by influenza A virus via the miRNA mechanism. The differential miRNA expressions can reveal the host-pathogen

Table 1 Common miRNAs expression profiles in response to influenza A virus infection obtained from several studies

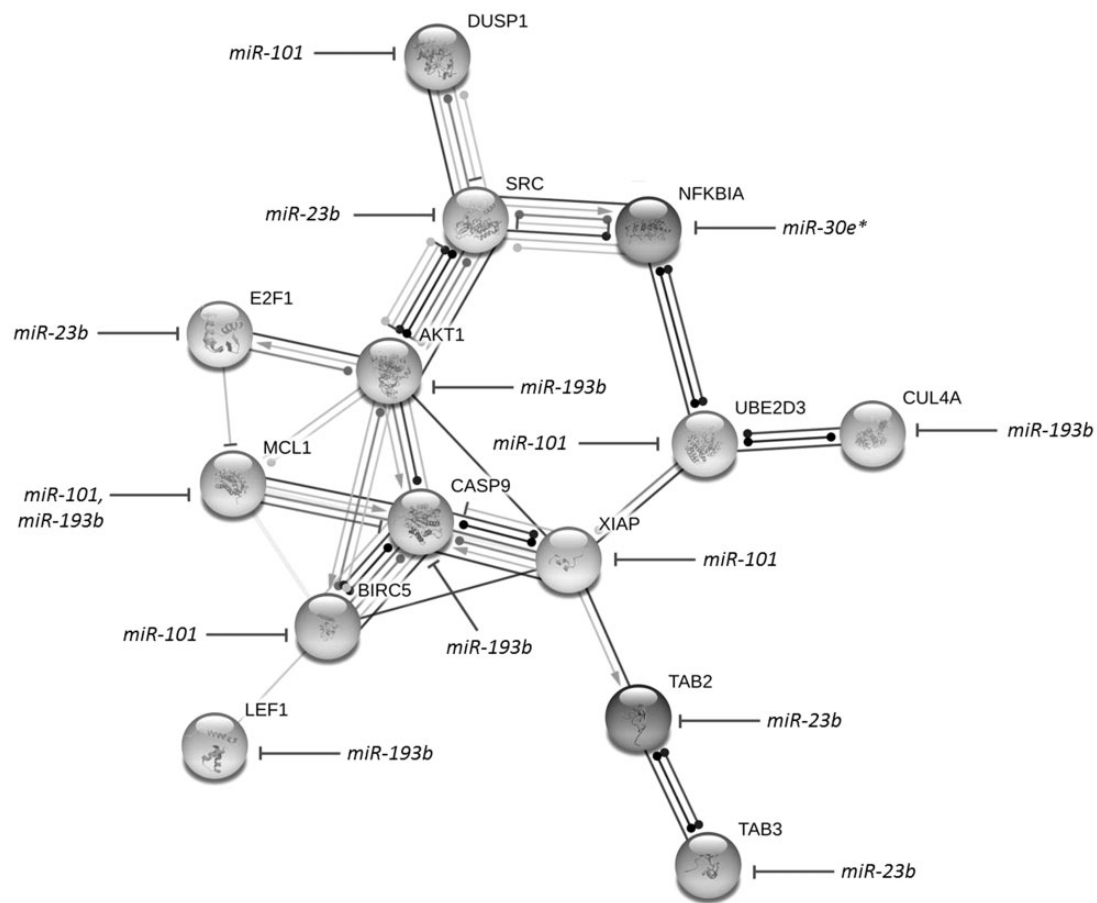
Study description		Differentially expressed miRNAs (Δ = Up regulated; ▽ = Down regulated)																	References
		Cells/ tissues	Viral subtype	Infection time	Methods	let-7	miR- 10	miR- 15	miR- 21	miR- 29	miR- 30	miR- 34	miR- 101	miR- 132	miR- 146	miR- 148	miR- 548	miR- 574	
Human	A549	pH1N1, H3N2, H5N1	24 h	NGS	Δ	▽	Δ	Δ	Δ	Δ	Δ	Δ	Δ	Δ	▽	Δ	Δ	Δ	This study
Human	A549	pH1N1	0, 4, 8, 24, 48, 72 h	qPCR	▽		▽	▽	▽	Δ									9
Human	A549	H1N1, H3N2	24 h	qPCR						Δ									31
Human	A549	H1N1, H3N2	4, 24 h	Array		Δ		Δ							Δ				41
Human	A549	H1N1, H3N2	3 h	Array & qPCR												▽			42
Human	A549	H1N1, H3N2	8, 12, 24 h	Array & qPCR						Δ				Δ	Δ				40
Human	BEAS-2B	H1N1, H3N2	8, 12, 24 h	Array & qPCR	▽	Δ								Δ	Δ			Δ	40
Human	NCI-H292	H1N1, H5N1	3 h	Array & qPCR					Δ									Δ	12
Macaques	Lungs	H5N1, pH1N1 (1918)	1, 2, 4, 7 d	Array & qPCR	▽	▽	Δ	Δ	Δ	▽	▽	▽	▽						28
Mice	Lungs	H5N1, pH1N1 (1918)	1, 3, 5 d	Array & qPCR	▽	▽	Δ	Δ	Δ	▽	▽	▽	▽						28
Mice	Lungs	H1N1	7 & 15 d	Array & qPCR	▽		Δ	Δ	Δ	▽	▽	▽							44
Dog	Lungs	H3N2	4 d	NGS & qPCR	Δ							Δ			▽				45
Pig	Lungs	H1N1, H3N2, H1N2	24 h, 72 h, 14 d	qPCR				Δ	Δ						▽				43

**Table 2** Numbers of validated target genes of common miRNAs in response to influenza A virus

miRNA family	Number and percentage of miRNA target genes based on GO biological process classification <sup>a</sup>													Total target genes
	AP	BA	BG	BR	CP	DP	IP	Lo	MO	MP	Re	Rs		
let-7	13	9	5	35	51	25	8	13	13	51	8	9	240	
	5.42%	3.75%	2.08%	14.58%	21.25%	10.42%	3.33%	5.42%	5.42%	21.25%	3.33%	3.75%		
miR-10	3	3	2	13	10	14	7	1	5	17	2	5	82	
	3.66%	3.66%	2.44%	15.85%	12.20%	17.07%	8.54%	1.22%	6.10%	20.73%	2.44%	6.10%		
miR-15	5	1	0	15	18	8	2	3	2	16	4	5	79	
	6.33%	1.27%	0.00%	18.99%	22.78%	10.13%	2.53%	3.80%	2.53%	20.25%	5.06%	6.33%		
miR-17	5	4	1	16	29	10	6	4	0	26	5	8	114	
	4.39%	3.51%	0.88%	14.04%	25.44%	8.77%	5.26%	3.51%	0.00%	22.81%	4.39%	7.02%		
miR-20	2	1	1	15	22	7	5	2	1	23	4	6	89	
	2.25%	1.12%	1.12%	16.85%	24.72%	7.87%	5.62%	2.25%	1.12%	25.84%	4.49%	6.74%		
miR-21	9	8	7	29	48	20	11	11	7	47	4	14	215	
	4.19%	3.72%	3.26%	13.49%	22.33%	9.30%	5.12%	5.12%	3.26%	21.86%	1.86%	6.51%		
miR-29	2	11	7	12	20	14	7	8	9	11	2	10	113	
	1.77%	9.73%	6.19%	10.62%	17.70%	12.39%	6.19%	7.08%	7.96%	9.73%	1.77%	8.85%		
miR-30	8	2	8	17	26	16	4	6	3	27	1	6	124	
	6.45%	1.61%	6.45%	13.71%	20.97%	12.90%	3.23%	4.84%	2.42%	21.77%	0.81%	4.84%		
miR-34	2	2	0	10	10	6	3	0	3	11	2	1	50	
	4.00%	4.00%	0.00%	20.00%	20.00%	12.00%	6.00%	0.00%	6.00%	22.00%	4.00%	2.00%		
miR-101	5	0	3	5	9	6	2	3	3	11	1	1	49	
	10.20%	0.00%	6.12%	10.20%	18.37%	12.24%	4.08%	6.12%	6.12%	22.45%	2.04%	2.04%		
miR-132	0	0	2	8	9	1	0	0	0	8	2	1	31	
	0.00%	0.00%	6.45%	25.81%	29.03%	3.23%	0.00%	0.00%	0.00%	25.81%	6.45%	3.23%		
miR-146	6	6	3	7	21	9	7	2	3	16	1	5	86	
	6.98%	6.98%	3.49%	8.14%	24.42%	10.47%	8.14%	2.33%	3.49%	18.60%	1.16%	5.81%		
miR-148	1	3	1	3	10	5	3	2	2	8	1	3	42	
	2.38%	7.14%	2.38%	7.14%	23.81%	11.90%	7.14%	4.76%	4.76%	19.05%	2.38%	7.14%		

Note: GO biological process of validated target genes was classified by PANTHER 9.0 classification system (<http://www.pantherdb.org/>) as the following: AP: apoptotic process (GO:0006915); BA: biological adhesion (GO:0022610); BG: biogenesis (GO:0071840); BR: biological regulation (GO:0065007); CP: cellular process (GO:0009987); DP: developmental process (GO:0032502); IP: immune system process (GO:0002378); Lo: localization (GO:0051179); MO: multicellular organismal process (GO:0032501); MP: metabolic process (GO:0008152); Re: reproduction (GO:0000003); Rs: response to stimulus (GO:0050896).

<sup>a</sup>Numbers of validated target genes were analyzed based on data (September 30, 2014) obtained from miRtarBase (<http://mirtarbase.mbc.nctu.edu.tw/>).



**Figure 3** MicroRNA regulated network of target genes related to apoptosis in response to influenza virus infection. The STRING 9.1, together with data from Panther 9.0 constructed the interactions between proteins playing an important role in programmed cell death. As integrated with *in silico* analysis of microRNA target prediction, the pathway revealed the differentially expressed microRNAs controlling numerous target genes

**Table 3** Viral targets of differentially expressed miRNAs in A549 infected with 3 subtypes of influenza virus

miRNAs	Ratios of infected/ non-infected (normalized)	Influenza virus segment
hsa-miR-127-3p	+1.35 (H3)	PB1
hsa-miR-128	+2.35 (H3) +1.96 (H5)	PA
hsa-miR-29a	+9.25 (H1) +1.74 (H5)	HA
hsa-miR-548	+1.15 (H5)	PB1, NS
hsa-miR-660	+1.74 (H5)	PA
hsa-miR-323	+1.62 (H3)	PB1
hsa-miR-let-7	+2.27 (H1)	M
hsa-miR-222	+2.22 (H5) +2.22 (H5)	PB2, PB1,PA,NS1, NP, M
hsa-miR-16	+2.97 (H5)	PB2, PB1,PA,NS1, NP, M
hsa-miR-136	-0.38 (H1)	NP, HA
hsa-miR-92a	-1.55 (H3)	PB2

interaction in viral infection in two ways. First, the virus can hijack the host cellular mechanism by up- or downregulating the miRNAs involved in pathways that can promote viral replication. For example, the results showed that

miR-181c was found to be upregulated in cells infected with H5N1 and H3N2 influenza A virus. The increase of this miRNA can target many genes associated with cellular immune defense, such as BCL2, IL-2, and Tnf,<sup>49-51</sup> and the deregulation of target genes can facilitate the influenza virus to avoid the suppressed host immune system. Second, the host cell can respond to viral infection via the miRNA mechanism by alteration of miRNAs which can have the effects on viral replication process to limit viral propagation. For example, the upregulation of miR-301 in H5N1 and H3N2 virus infection which can inhibit NKRF, the NFKB repressing factor, will promote the cytokine which indicates the innate and adaptive immunity response in mammalian cells.<sup>52</sup>

Many previous studies have suggested a correlation between influenza A virus infection and miRNA alteration. The infection of influenza virus can specifically activate the alteration of miRNAs including let-7 family, miR-30a-c, miR-132 and miR-30e\* which might target in many genes function in lung repair.<sup>44,53</sup> However, the previous study found that miRNAs in let-7 family (let-7c, -7f, -7i), miR-30b and miR-26b were also upregulated by respiratory syncytial virus (RSV) infection,<sup>53</sup> implying that the alteration of these miRNAs are not only present against influenza virus infection but also other respiratory viruses.



The miRNA expression profiles were analyzed and compared to those from several studies which were obtained from different sample sources, viral strains, infection periods, and culture models, in order to discover the common miRNAs that overlapped between different systems. The results showed that some miRNAs were required to be considered. The miR-21, -29, -30, -34 and let-7 family were widely responsive to influenza virus infection in many systems. Interestingly, miR-15 and miR-21 were upregulated by influenza virus infection while miR-132, -30, and -101 were upregulated only in the human model system.

After the target prediction had been performed, most of the mechanisms due to virus infection were related to cellular processes, metabolic processes, developmental processes, and biological regulation (Table 2 and Table S1, Supplementary Information). Many miRNAs and their target genes should be considered and further discussed. MiR-101 can target MCL1, the anti-apoptosis gene and PRKDC, one of the large family of serine/threonine kinase,<sup>54</sup> which can play an important role in the receptor-mediated endocytosis process of the influenza life cycle, since the HA protein of the influenza virus can efficiently activate the PKC signaling pathway.<sup>55,56</sup> It has been reported that the influenza virus propagation process can be suppressed by a specific inhibitor of PKC.<sup>57</sup> Another miRNA that should be considered is the regulation of cell apoptosis via miR-29c (Figure 3) which was found to be upregulated in H5N1 infection. Much evidence suggests that this miRNA is involved in the virus-induced apoptosis pathway via the repression of anti-apoptotic agent, BCL2L2.<sup>41</sup> This is related to a previous study that found that the miR-29 family tends to be more upregulated due to influenza virus infection.<sup>58</sup> Moreover, some of the miRNA experimentally targeted genes play a critical role in immunoregulatory functions. For example, the upregulation of miR-15b suggests that infection of influenza virus can suppress the host immune response via miRNA mechanism. A previous study found that the IL-1B gene, the crucial regulator of inflammatory response, cellular activity including cell proliferation, differentiation and apoptosis, can decrease the expression of miR-let-7g and miR-26b.<sup>53</sup> Corresponding to this study, the high expression of let-7g and miR-26b indicated that the IL-1B has not been produced in the first 24h after infection. In addition, our study showed that the upregulation of let-7 family (in all 3 subtypes infection), miR-29a, b (in pH1N1 infection) and miR-29c (in H5N1 infection) might have resulted from TNF- $\alpha$  and IL-4 still being inactive in the early phase of infection. However, the downregulation of miR-26b by H3N2 influenza virus infection should be considered, as it might be responsible for the different pathogenicity of H3N2 virus from pH1N1 virus and RSV.

A number of miRNAs have been found to be down regulated. Most of them can target genes associated with DNA methylation, protein degradation, and the signal transduction pathway. For example, the downregulation of miR-125b can trigger the MAPK signaling pathway, which regulates various cellular responses such as cell proliferation and apoptosis.<sup>59,60</sup> Previously, MAPK and CDK13 can be controlled by miR-548-3p and miR-138, which were,

respectively,<sup>61</sup> identified as the mediators required for influenza virus replication. It is tempting to conclude that the upregulation of miR-548 (in H5N1 infection) and miR-138 (in pH1N1 infection) is the mechanism of the host response to limit influenza virus replication. Besides, a previous study suggested that the upregulation of MAPK protein can result from HA membrane accumulation due to the increase of polymerase activity. Interestingly, the upregulation of genes associated with DNA methylation via the decline of miRNA can cause the chromatin packaging, which can lead to the protein expression inhibition and may affect virus replication.

The difference of miRNA expression profile in cells infected with different subtypes might provide insight in the distinct pathogenicity of influenza virus. The result showed that there are four upregulated miRNAs including hsa-miR-101, hsa-miR-193b, hsa-miR-23b, and hsa-miR-30e\*. Moreover, there were some strain-specific miRNAs which expressed in different levels in each subtype of influenza virus. The results showed that infection of H5N1 HPAI influenza virus can cause the highest amount of differential miRNA expression. Compared to a previous study, miR-31 and miR-29c which are upregulated in H5N1 infection, and miR-29a and b, which are upregulated in pH1N1 infection, were confirmed to be potentially controlled during the early stage of influenza virus infection.<sup>38</sup> Interestingly, the results showed that miR-484 was found to be deregulated in cells infected with H5N1 and H3N2 influenza A virus. This miRNA can inhibit furin, the proteolytic enzyme that can cleave HA0 protein to facilitate influenza virus entry step. The downregulation of miR-484 will allow the host cell to produce more furin and promote the furin-dependent entry mechanism of the influenza viral life cycle.<sup>62-64</sup> One point of discussion should be considered, infection of the pH1N1 virus can recruit the miR-30 family, including miR-30a, miR-30a\*, miR-30b, and miR-30c while only one member of miR-30 family was found to be deregulated in H3N2 infection, contrary to a previous study.<sup>11</sup> However, the previous study suggested that genetics of the subject might account for the regulation of miRNA expression,<sup>65</sup> so that genetic variation and a different cell culture system can affect the miRNA expression during influenza virus infection. While the previous study found that the change of miR-21\*, miR-100\*, miR-141, miR-574-3p, miR-1274a and b resulted from H5N1 infection in NCI-H292 cell line,<sup>12</sup> our results show the upregulation of miR-21\* only. This suggested that the different cell culture might be responsible for various cellular responses via the miRNA mechanism. Moreover, compared to a previous study,<sup>66</sup> which investigated the miRNA profile of A/Udorn/72 (H3N2) infection, and suggested that the miRNA profile of the same subtype of influenza can be altered depending on the different genetic characteristics of the virus, the up-to-date virus infection can represent the virus that is presently circulating. In the study of Zhao et al.,<sup>45</sup> infection of canine influenza virus in lungs and trachea of dogs can cause the abundant expression of miR-143 and let-7. When compared to our study, the difference of miR-143 expression level may be due to the different characteristics of viral strains or host cells.

Infection with virus can alter many cellular mechanisms such as apoptosis which is one of the main machinery. Herein, we identified the genes and miRNAs observed in high-throughput sequencing that are involved in the apoptotic pathway by using STRING software (Figure 3). The pathway revealed the differentially expressed miRNAs controlling numerous target genes including NFkB, SOC, STAT, and IFN which can be regulated by many interesting miRNAs, including let-7 family, miR-101, miR-30, and miR-21, which might lead to development of miRNA therapeutics for influenza-like illness. However, many further experiments should be required such as western blot, *in vivo* study and pull down assays to make the refined observation and development.

Most of the upregulated miRNAs were predicted to target the influenza virus genome (Table 3). Most of them could bind to genes that have been reported in the early stage of influenza virus life cycle such as hemagglutinin and polymerase (PB2, PB1 and PA), which play the vital role in influenza virus entry mechanism and genome replication, respectively. In addition, a previous study suggested that miR-548an can mediate the influenza virus infection by regulation of non-structural-1A binding protein (NS1ABP),<sup>42</sup> corresponding to our study which found that the miR-548 was upregulated in H5N1 infection, implying that the host cell can produce more miR-548 against the exposure of influenza virus from an early stage of infection. Some miRNAs were found to be downregulated, suggesting that the virus can also trigger the miRNA pathway to fine-tune the miRNA mechanism to survive in host cells. The study in other time points after virus infection might reveal other differential miRNAs that can target other sites of the influenza virus genome, which may lead to the insight into the cellular mechanisms used to fight back against influenza virus infection. However, the built-on study by using widely used panel of robust tools, which are specific miRNAs inhibitors and miRNA mimics to knock down or increase the expression of miRNAs, is crucial to investigate the specific role of these candidate miRNAs.

## Conclusion

In summary, this study provides the overall analysis of miRNA profiles in cell cultures infected with three different subtypes of influenza virus which have caused sporadic outbreaks in the human population, to gain insight into the overview of how the influenza virus can trigger the host cellular mechanism to facilitate itself to survive in host cells and also to find how cells can manipulate the cellular process to defend against influenza virus infection via the microRNA mechanism. This study provides information only in the early stage of infection, at 24 h post infection and can generate some potential candidate miRNAs and predicted target genes. This information can lead to the development of effective therapeutic agents, which have to interfere or control the influenza life cycle at an early stage of infection. Moreover, to explore the novel antiviral drugs, further analysis will be required such as miRNA profiling in late stages of infection or the study of knock-down and overexpression of candidate miRNAs.

**Authors' contributions:** JM analyzed the miRNA profiling and drafted the manuscript. WP carried out the NGS process, data processing and visualization. SS performed protein interaction network. KK assisted in cell culture and virus infection. KP assisted in NGS libraries preparation. TJ assisted in data analysis. YP provided stock of viruses and revised the manuscript. SP designed the study, data analysis, revised the manuscript and coordination. All authors read and approved the final manuscript.

## ACKNOWLEDGEMENTS

The authors would like to gratefully acknowledge the Department of Biochemistry and Research affairs, Faculty of Medicine, Chulalongkorn University for their invaluable contribution to the success of this study. Funding for this study was supported by the Thailand Research Fund (TRF: RSA5680031); the Postdoctoral Scholarship, Ratchadapiseksompotch Fund (Faculty of Medicine, Chulalongkorn University); National Research University Project; Office of Higher Education Commission (WCU-007-HR-57); Centenary Academic Development Project (CU56-HR01); the Ratchadaphiseksomphot Endowment Fund of Chulalongkorn University (RES560530093); Research Chair Grant, National Science and Technology Development Agency (NSTDA) and Chulalongkorn Academic Advancement into its 2nd Century Project.

## DECLARATION OF CONFLICTING INTERESTS

The authors hereby declare no personal or professional conflicts of interest with any aspect of this study.

## REFERENCES

- Baum M, Bielau S, Rittner N, Schmid K, Eggelbusch K, Dahms M, Schlauersbach A, Tahedl H, Beier M, Guimil R, Scheffler M, Hermann C, Funk JM, Wixmerten A, Rebscher H, Honig M, Andrae C, Buchner D, Moschel E, Glathe A, Jager E, Thom M, Greil A, Bestvater F, Obermeier F, Burgmaier J, Thome K, Weichert S, Hein S, Binnewies T, Foitzik V, Muller M, Stahler CF, Stahler PF. Validation of a novel, fully integrated and flexible microarray benchtop facility for gene expression profiling. *Nucleic Acids Res* 2003;**31**:e151
- Smith GJ, Vijaykrishna D, Bahl J, Lycett SJ, Worobey M, Pybus OG, Ma SK, Cheung CL, Raghwani J, Bhatt S, Peiris JS, Guan Y, Rambaut A. Origins and evolutionary genomics of the 2009 swine-origin H1N1 influenza A epidemic. *Nature* 2009;**459**:1122–5
- Dawood FS, Iuliano AD, Reed C, Meltzer MI, Shay DK, Cheng PY, Bandaranayake D, Breiman RF, Brooks WA, Buchy P, Feikin DR, Fowler KB, Gordon A, Hien NT, Horby P, Huang QS, Katz MA, Krishnan A, Lal R, Montgomery JM, Molbak K, Pebody R, Presanis AM, Razuri H, Steens A, Tinoco YO, Wallinga J, Yu H, Vong S, Bresee J, Widdowson MA. Estimated global mortality associated with the first 12 months of 2009 pandemic influenza A H1N1 virus circulation: a modelling study. *Lancet Infect Dis* 2012;**12**:687–95
- Yuen KY, Chan PK, Peiris M, Tsang DN, Que TL, Shortridge KF, Cheung PT, To WK, Ho ET, Sung R, Cheng AF. Clinical features and rapid viral diagnosis of human disease associated with avian influenza A H5N1 virus. *Lancet* 1998;**351**:467–71
- Tran TH, Nguyen TL, Nguyen TD, Luong TS, Pham PM, Nguyen v V, Pham TS, Vo CD, Le TQ, Ngo TT, Dao BK, Le PP, Nguyen TT, Hoang TL, Cao VT, Le TG, Nguyen DT, Le HN, Nguyen KT, Le HS, Le VT, Christiane D, Tran TT, Menno de J, Schultz C, Cheng P, Lim W, Horby P, Farrar J, World Health Organization International Avian Influenza Investigative T. Avian influenza A (H5N1) in 10 patients in Vietnam. *New Engl J Med* 2004;**350**:1179–88

6. Chan PK. Outbreak of avian influenza A(H5N1) virus infection in Hong Kong in 1997. *Clin Infect Dis: an official publication of the Infectious Diseases Society of America* 2002;**34**:S58–64
7. Peiris JS, Yu WC, Leung CW, Cheung CY, Ng WF, Nicholls JM, Ng TK, Chan KH, Lai ST, Lim WL, Yuen KY, Guan Y. Re-emergence of fatal human influenza A subtype H5N1 disease. *Lancet* 2004;**363**:617–9
8. Komar N, Olsen B. Avian influenza virus (H5N1) mortality surveillance. *Emerg Infect Dis* 2008;**14**:1176–8
9. Loveday EK, Svinti V, Diederich S, Pasick J, Jean F. Temporal- and strain-specific host microRNA molecular signatures associated with swine-origin H1N1 and avian-origin H7N7 influenza A virus infection. *J Virol* 2012;**86**:6109–22
10. Meliopoulos VA, Andersen LE, Birrer KF, Simpson KJ, Lowenthal JW, Bean AG, Stambas J, Stewart CR, Tompkins SM, van Beusechem VW, Fraser I, Mhlanga M, Barichievy S, Smith Q, Leake D, Karpilow J, Buck A, Jona G, Tripp RA. Host gene targets for novel influenza therapies elucidated by high-throughput RNA interference screens. *FASEB J: official publication of the Federation of American Societies for Experimental Biology* 2012;**26**:1372–86
11. Wang Y, Brahmakshatriya V, Lupiani B, Reddy SM, Soibam B, Benham AL, Gunaratne P, Liu HC, Trakooljul N, Ing N, Okimoto R, Zhou H. Integrated analysis of microRNA expression and mRNA transcriptome in lungs of avian influenza virus infected broilers. *BMC Genomics* 2012;**13**:278
12. Lam WY, Yeung AC, Ngai KL, Li MS, To KF, Tsui SK, Chan PK. Effect of avian influenza A H5N1 infection on the expression of microRNA-141 in human respiratory epithelial cells. *BMC Microbiol* 2013;**13**:104
13. Wu Z, Hao R, Li P, Zhang X, Liu N, Qiu S, Wang L, Wang Y, Xue W, Liu K, Yang G, Cui J, Zhang C, Song H. MicroRNA expression profile of mouse lung infected with 2009 pandemic H1N1 influenza virus. *PloS ONE* 2013;**8**:e74190
14. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;**116**:281–97
15. Lagos-Quintana M, Rauhut R, Lendeckel W, Tuschl T. Identification of novel genes coding for small expressed RNAs. *Science* 2001;**294**:853–8
16. Lau NC, Lim LP, Weinstein EG, Bartel DP. An abundant class of tiny RNAs with probable regulatory roles in *Caenorhabditis elegans*. *Science* 2001;**294**:858–62
17. Lee RC, Ambros V. An extensive class of small RNAs in *Caenorhabditis elegans*. *Science* 2001;**294**:862–4
18. Ambros V. MicroRNAs and developmental timing. *Curr Opin Genet Develop* 2011;**21**:511–7
19. Lee RC, Feinbaum RL, Ambros V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell* 1993;**75**:843–54
20. Griffiths-Jones S. The microRNA Registry. *Nucl Acids Res* 2004;**32**:D109–11
21. Griffiths-Jones S, Grocock RJ, van Dongen S, Bateman A, Enright AJ. miRBase: microRNA sequences, targets and gene nomenclature. *Nucleic Acids Res* 2006;**34**:D140–4
22. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, Shimizu M, Rattan S, Bullrich F, Negrini M, Croce CM. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci USA* 2004;**101**:2999–3004
23. John B, Enright AJ, Aravin A, Tuschl T, Sander C, Marks DS. Human microRNA targets. *PLoS Biol* 2004;**2**:e363
24. Wang B, Koh P, Winbanks C, Coughlan MT, McClelland A, Watson A, Jandeleit-Dahm K, Burns WC, Thomas MC, Cooper ME, Kantharidis P. miR-200a Prevents renal fibrogenesis through repression of TGF-beta2 expression. *Diabetes* 2011;**60**:280–7
25. Gottwein E, Cullen BR. Viral and cellular microRNAs as determinants of viral pathogenesis and immunity. *Cell Host Microbe* 2008;**3**:375–87
26. Skalsky RL, Cullen BR. Viruses, microRNAs, and host interactions. *Ann Rev Microbiol* 2010;**64**:123–41
27. Li Y, Chan EY, Li J, Ni C, Peng X, Rosenzweig E, Tumpey TM, Katze MG. MicroRNA expression and virulence in pandemic influenza virus-infected mice. *J Virol* 2010;**84**:3023–32
28. Li Y, Li J, Belisle S, Baskin CR, Tumpey TM, Katze MG. Differential microRNA expression and virulence of avian, 1918 reassortant, and reconstructed 1918 influenza A viruses. *Virology* 2011;**421**:105–13
29. Rogers JV, Price JA, Wendling MQ, Long JP, Bresler HS. Preliminary microRNA analysis in lung tissue to identify potential therapeutic targets against H5N1 infection. *Viral Immunol* 2012;**25**:3–11
30. Song L, Liu H, Gao S, Jiang W, Huang W. Cellular microRNAs inhibit replication of the H1N1 influenza A virus in infected cells. *J Virol* 2010;**84**:8849–60
31. Terrier O, Textoris J, Carron C, Marcel V, Bourdon JC, Rosa-Calatrava M. Host microRNA molecular signatures associated with human H1N1 and H3N2 influenza A viruses reveal an unanticipated antiviral activity for miR-146a. *J General Virol* 2013;**94**:985–95
32. Geiss GK, An MC, Bumgarner RE, Hammersmark E, Cunningham D, Katze MG. Global impact of influenza virus on cellular pathways is mediated by both replication-dependent and -independent events. *J Virol* 2001;**75**:4321–31
33. Severson WE, McDowell M, Ananthan S, Chung DH, Rasmussen L, Sosa MI, White EL, Noah J, Jonsson CB. High-throughput screening of a 100,000-compound library for inhibitors of influenza A virus (H3N2). *J Biomol Screen* 2008;**13**:879–87
34. Shih SR, Chu TY, Reddy GR, Tseng SN, Chen HL, Tang WF, Wu MS, Yeh JY, Chao YS, Hsu JT, Hsieh HP, Horng JT. Pyrazole compound BPR1P0034 with potent and selective anti-influenza virus activity. *J Biomed Sci* 2010;**17**:13
35. Borjabad A, Morgello S, Chao W, Kim SY, Brooks AI, Murray J, Potash MJ, Volsky DJ. Significant effects of antiretroviral therapy on global gene expression in brain tissues of patients with HIV-1-associated neurocognitive disorders. *PLoS Pathogen* 2011;**7**:e1002213
36. Mi H, Muruganujan A, Thomas PD. PANTHER in 2013: modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucl Acid Res* 2013;**41**:D377–86
37. Szklarczyk D, Franceschini A, Kuhn M, Simonovic M, Roth A, Minguez P, Doerks T, Stark M, Muller J, Bork P, Jensen LJ, von Mering C. The STRING database in 2011: functional interaction networks of proteins, globally integrated and scored. *Nucl Acid Res* 2011;**39**:D561–8
38. Song H, Wang Q, Guo Y, Liu S, Song R, Gao X, Dai L, Li B, Zhang D, Cheng J. Microarray analysis of microRNA expression in peripheral blood mononuclear cells of critically ill patients with influenza A (H1N1). *BMC Infect Dis* 2013;**13**:257
39. von Mering C, Jensen LJ, Snel B, Hooper SD, Krupp M, Foglierini M, Jouffre N, Huynen MA, Bork P. STRING: known and predicted protein-protein associations, integrated and transferred across organisms. *Nucl Acid Res* 2005;**33**:D433–7
40. Buggele WA, Johnson KE, Horvath CM. Influenza A virus infection of human respiratory cells induces primary microRNA expression. *J Biol Chem* 2012;**287**:31027–40
41. Guan Z, Shi N, Song Y, Zhang X, Zhang M, Duan M. Induction of the cellular microRNA-29c by influenza virus contributes to virus-mediated apoptosis through repression of antiapoptotic factors BCL2L2. *Biochem Biophys Res Commun* 2012;**425**:662–7
42. Othumpangat S, Noti JD, Blachere FM, Beezhold DH. Expression of non-structural-1A binding protein in lung epithelial cells is modulated by miRNA-548an on exposure to influenza A virus. *Virology* 2013;**447**:84–94
43. Skovgaard K, Cirera S, Vasby D, Podolska A, Breum SO, Durrwald R, Schlegel M, Heegaard PM. Expression of innate immune genes, proteins and microRNAs in lung tissue of pigs infected experimentally with influenza virus (H1N2). *Innate Immun* 2013;**19**:531–44
44. Tan KS, Choi H, Jiang X, Yin L, Seet JE, Patzel V, Engelward BP, Chow VT. Micro-RNAs in regenerating lungs: an integrative systems biology analysis of murine influenza pneumonia. *BMC Genom* 2014;**15**:587
45. Zhao FR, Su S, Zhou DH, Zhou P, Xu TC, Zhang LQ, Cao N, Qi WB, Zhang GH, Li SJ. Comparative analysis of microRNAs from the lungs and trachea of dogs (*Canis familiaris*) infected with canine influenza virus. *Infect Genet Evolut: journal of molecular epidemiology and evolutionary genetics in infectious diseases* 2014;**21**:367–74



46. Lecellier CH, Dunoyer P, Arar K, Lehmann-Che J, Eyquem S, Himber C, Saib A, Voinnet O. A cellular microRNA mediates antiviral defense in human cells. *Science* 2005;**308**:557–60
47. Scaria V, Hariharan M, Pillai B, Maiti S, Brahmachari SK. Host-virus genome interactions: Macro roles for microRNAs. *Cell Microbiol* 2007;**9**:2784–94
48. Khongnomnan K, Poomipak W, Poovorawan Y, Payungporn S. In silico Analysis of Human microRNAs Targeting Influenza A Viruses (subtype H1N1, H5N1 and H3N2). *World Acad Sci Eng Technol* 2012;**71**:433–9
49. Xue Q, Guo ZY, Li W, Wen WH, Meng YL, Jia LT, Wang J, Yao LB, Jin BQ, Wang T, Yang AG. Human activated CD4(+) T lymphocytes increase IL-2 expression by downregulating microRNA-181c. *Molecul Immunol* 2011;**48**:592–9
50. Ouyang YB, Lu Y, Yue S, Giffard RG. miR-181 targets multiple Bcl-2 family members and influences apoptosis and mitochondrial function in astrocytes. *Mitochondrion* 2012;**12**:213–9
51. Sun X, Icli B, Wara AK, Belkin N, He S, Kobzik L, Hunninghake GM, Vera MP, Registry M, Blackwell TS, Baron RM, Feinberg MW. MicroRNA-181b regulates NF-kappaB-mediated vascular inflammation. *J Clin Invest* 2012;**122**:1973–90
52. Chu WM, Ostertag D, Li ZW, Chang L, Chen Y, Hu Y, Williams B, Perrault J, Karin M. JNK2 and IKKbeta are required for activating the innate response to viral infection. *Immunity* 1999;**11**:721–31
53. Globinska A, Pawelczyk M, Kowalski ML. MicroRNAs and the immune response to respiratory virus infections. *Expert Rev Clin Immunol* 2014;**10**:963–71
54. Toker A. Signaling through protein kinase C. *Front Biosci: a journal and virtual library* 1998;**3**:D1134–47
55. Constantinescu SN, Cernescu CD, Popescu LM. Effects of protein kinase C inhibitors on viral entry and infectivity. *FEBS Lett* 1991;**292**:31–3
56. Kunzelmann K, Beesley AH, King NJ, Karupiah G, Young JA, Cook DI. Influenza virus inhibits amiloride-sensitive Na<sup>+</sup> channels in respiratory epithelia. *Proc Natl Acad Sci USA* 2000;**97**:10282–7
57. Root CN, Wills EG, McNair LL, Whittaker GR. Entry of influenza viruses into cells is inhibited by a highly specific protein kinase C inhibitor. *J General Virol* 2000;**81**:2697–705
58. Zhang X, Dong C, Sun X, Li Z, Zhang M, Guan Z, Duan M. Induction of the cellular miR-29c by influenza virus inhibits the innate immune response through protection of A20 mRNA. *Biochem Biophys Res Commun* 2014;**450**:755–61
59. Pearson G, Robinson F, Beers Gibson T, Xu BE, Karandikar M, Berman K, Cobb MH. Mitogen-activated protein (MAP) kinase pathways: regulation and physiological functions. *Endocrine Rev* 2001;**22**:153–83
60. Dong C, Davis RJ, Flavell RA. MAP kinases in the immune response. *Ann Rev Immunol* 2002;**20**:55–72
61. Bakre A, Andersen LE, Meliopoulos V, Coleman K, Yan X, Brooks P, Crabtree J, Tompkins SM, Tripp RA. Identification of host kinase genes required for influenza virus replication and the regulatory role of microRNAs. *PLoS ONE* 2013;**8**:e66796
62. Stienke-Grober A, Vey M, Angliker H, Shaw E, Thomas G, Roberts C, Klenk HD, Garten W. Influenza virus hemagglutinin with multibasic cleavage site is activated by furin, a subtilisin-like endoprotease. *EMBO J* 1992;**11**:2407–14
63. Horimoto T, Nakayama K, Smeekens SP, Kawaoka Y. Proprotein-processing endoproteases PC6 and furin both activate hemagglutinin of virulent avian influenza viruses. *J Virol* 1994;**68**:6074–8
64. Luna C, Li G, Qiu J, Epstein DL, Gonzalez P. MicroRNA-24 regulates the processing of latent TGFbeta1 during cyclic mechanical stress in human trabecular meshwork cells through direct targeting of FURIN. *J Cell Physiol* 2011;**226**:1407–14
65. Bueno MJ, Perez de Castro I, Gomez de Cedron M, Santos J, Calin GA, Cigudosa JC, Croce CM, Fernandez-Piqueras J, Malumbres M. Genetic and epigenetic silencing of microRNA-203 enhances ABL1 and BCR-ABL1 oncogene expression. *Cancer cell* 2008;**13**:496–506
66. Buggele WA, Krause KE, Horvath CM. Small RNA profiling of influenza A virus-infected cells identifies miR-449b as a regulator of histone deacetylase 1 and interferon beta. *PLoS ONE* 2013;**8**:e76560

(Received May 20, 2015, Accepted September 21, 2015)